



An alternative dogma on reduced artemisinin susceptibility: A new shadow from east to west

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In PNAS, Demas et al. (1) show, by long-term in vitro selection using culture-adapted *Plasmodium falciparum* isolates from Senegal, that the gene encoding the actin-binding protein *P. falciparum* coronin (*pfcoronin*) and its genetic variants (G50E, R100K, and E107V) can reduce the susceptibility of the parasite to the active metabolite of the fast-acting antimalarial drug artemisinin, dihydroartemisinin (DHA). Resistance to artemisinins is a global threat in malaria control and elimination efforts (2).

Artemisinin resistance, first reported in Southeast Asia and still extremely rare, was associated with the *P. falciparum* PfKelch13-propeller domain (*kelch13* mutations: Y493H, R539T, I543T, and C580Y) (3). PfCoronin, which is structurally similar to Kelch13, is believed to interact with F-actin via its N-terminal propeller domain and to mediate actin organization and motility in merozoites and sporozoites (4, 5). The worldwide map of the occurrence of *kelch13*, however, indicates absence of the Asian artemisinin-resistance alleles in Africa (6–8). So far, it is not clear whether the *pfcoronin* variants G50E, R100K, and E107V occur in natural *P. falciparum* populations—in particular, in clinical isolates from Africa.

We looked at a total of 353 *P. falciparum* patient isolates that were earlier characterized for the absence of *kelch13* gene mutations (7–10) from 4 African countries to verify whether these isolates carry the *pfcoronin* mutations G50E, R100K, and E107V, which were described by Demas et al. (1) to be associated with reduced susceptibility to DHA. A total of 297 samples

were successfully genotyped by direct Sanger sequencing. Details of the study groups from Gabon ($n = 102$), Congo ($n = 48$), Ghana ($n = 57$), and Kenya ($n = 90$) are described elsewhere (7–10). The *pfcoronin* mutations G50E, R100K, and E107V were not observed at all among the isolates. However, 14 distinct mutations, including several nonsynonymous substitutions, were identified in the *pfcoronin* exon-3 (Table 1). None of the isolates carried the Asian *kelch13* resistance alleles M476I, Y493H, R539T, I543T, and C580Y. The mutation P76S (DNA position C562T) was observed to be most frequent (>10%) among isolates from central and west Africa. There was no indication of artemisinin or artemisinin-based combination therapy resistance in these patients. The functional role of the observed *pfcoronin* P76S needs to be elucidated among central and west African *P. falciparum* isolates.

Much effort has been made in recent years to determine the genetic basis of artemisinin resistance, which still remains unclear to a large extent. There is an obvious difference in occurrence of *pfkelch13* and *pfcoronin* alleles between Asia and Africa, which may also cause differences in parasite clearance rates during treatment with artemisinin-containing antimalarial combinations. However, we should bear in mind that parasite clearance rate or failure of an artemisinin-containing antimalarial is also, and even most often, determined by the activity of the partner drug, such as lumefantrine, amodiaquine, piperaquine, and pyronaridine.

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Table 1. pfcoronin mutations observed in 4 African countries

| Amino acid change | SNP position | Gabon, n (%) (n = 102) | Ghana, n (%) (n = 57) | Kenya, n (%) (n = 90) | Congo, n (%) (n = 48) | Total, n (%) (N = 297) |
|-------------------|--------------|---------------------------|--------------------------|--------------------------|--------------------------|---------------------------|
| I53I | C495T | 0 | 1 (2) | 0 | 0 | 1 (0.3) |
| V62M | G520A | 1 (1) | 1 (2) | 0 | 0 | 2 (0.7) |
| K69K/I/R | A542A/T/G | 0 | 3 (5) | 0 | 11 (23) | 14 (5) |
| P76S | C562T | 11 (11) | 9 (16) | 4 (4) | 8 (17) | 32 (11) |
| N110N/Y/D | A664A/T/G | 0 | 1 (2) | 0 | 5 (10) | 6 (2) |
| N112N/Y/D | A670A/T/G | 0 | 0 | 0 | 1 (2) | 1 (0.3) |
| K115K/stop/E | A679A/T/G | 0 | 0 | 0 | 1 (2) | 1 (0.3) |
| L121L/F/L | A699A/T/G | 0 | 0 | 0 | 1 (2) | 1 (0.3) |
| K127K/stop/E | A715A/T/G | 0 | 0 | 0 | 1 (2) | 1 (0.3) |
| K127K/I/R | A716A/T/G | 0 | 0 | 0 | 5 (10) | 5 (2) |
| V128V | A720A/T/G | 0 | 0 | 0 | 1 (2) | 1 (0.3) |
| N134N/Y/D | A736A/T/G | 0 | 0 | 0 | 4 (8) | 4 (1) |
| N137N/Y/D | A745A/T/G | 0 | 0 | 0 | 10 | 10 (3) |
| N137N/I/S | A746A/T/G | 0 | 0 | 0 | 2 (4) | 2 (0.7) |

SNP, single nucleotide polymorphism.

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